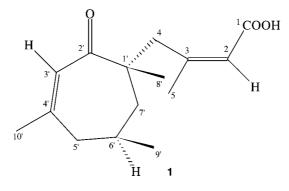
Leptosphaeric Acid, a Metabolite with a Novel Carbon Skeleton from Leptosphaeria sp. IV403, an Endophytic Fungus in Artemisia annua

by Jun-Yan Liu, Chang-Hong Liu, Wen-Xin Zou, Ren-Xiang Tan*

Institute of Functional Biomolecules, State Key Laboratory of Pharmaceutical Biotechnology, Nanjing University, Nanjing 210093, People's Republic of China (phone: +86-25-3592945; fax: +86-25-3302728; e-mail: rxtan@netra.nju.edu.cn)

Leptosphaeric acid (1), a metabolite with a novel carbon skeleton, was isolated from the AcOEt extract of the culture of *Leptosphaeria* sp., an endophytic fungus (strain number: IV403) of *Artemisia annua*. The structure of acid 1 was elucidated on the basis of spectral analyses including homo- and heteronuclear correlation NMR experiments (HMQC and HMBC). The absolute configuration of the metabolite was determined by its CD and NOED studies.

1. Introduction. – Endophytes, microorganisms living inside the healthy plant tissue, have been confirmed to be a rich source of functional biomolecules [1]. In the continuation of our characterization of chemically new and/or biologically active metabolites from the cultures of endophytic fungi of *Artemisia* species [2-4], another metabolite with a new carbon framework named leptosphaeric acid (1) was obtained from the AcOEt extract of the re-culture broth of *Leptosphaeria* sp., an endophytic fungus (strain number: IV403) of *Artemisia annua*. We herewith present the establishment of the structure and absolute configuration of compound **1**.



2. Results and Discussion. – Compound 1 was obtained as white gum. Its highresolution ESI mass spectrum (positive-ion mode) showed intense quasimolecular ions at m/z 251.1641 ($[M + H]^+$) and 273.1640 ($[M + Na]^+$), respectively, corresponding to the molecular formula $C_{15}H_{22}O_3$ (the index of unsaturation: 5), which was well consistent with a total of 15 discrete signals in its ¹³C-NMR spectrum edited by the DEPT pulse sequences. The presence of a conjugated COOH group and an α,β unsaturated ketone was implied by the UV peak at 242 nm, and the IR absorption bands at 3500-2500, 1713.3, 1692.9, 1668.4, and 1642.9 cm^{-1} , indicative of two C=O groups (C(1) and C(2')) and a pair of C=C bonds (between C(2) and C(3) as well as between C(3') and C(4')) disclosed also by the ¹³C-NMR spectrum. Subtraction of the four indexes of unsaturation involved in the above-mentioned moieties from that of the molecule demonstrated that compound **1** had to be monocyclic.

The conclusion were drawn on the basis of detailed NMR analyses (DEPT, HMQC, HMBC, and NOED) leading to the unambiguous assignments of ¹H- and ¹³C-NMR data (*Table*). The discerned HMBC correlations (*Table*), along with coupling constants and ¹³C-NMR data, established three substructures **A**, **B**, and **C** (the C-atoms were numbered in the same manner as for **1**). Specifically, the HMBC correlations of H-C(3') to C(5') and C(10'), of H-C(10') with C(3') and C(5'), and of H-C(5') to C(3') demonstrated the partial structure **A**, while substructure **B** was deduced from the HMBC correlations of H-C(2) to C(4) and C(5), and of H-C(5) to C(2) and C(4). Furthermore, the fragment **C** afforded by the sequential couplings from H-C(5') to H-C(7') (*Table*) was also reinforced by the HMBC correlations of H-C(7') with C(5'), of H-C(6') to C(1'), of H-C(9') to C(7'), and of H-C(8') with C(7'). The substructure **A** sharing a pivotal CH_2 (C(5')) with fragment **C** established the connection between the two subunits. In view of the imperative presence of a monocycle in the molecule, both C(2') in substructure **A** and C(4) in fragment **B** had to

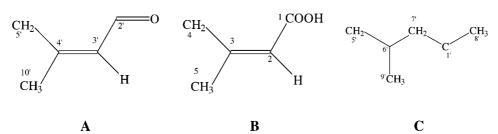


Table. ¹H- and ¹³C-NMR Data (CDCl₃) of Compound **1**

Position	$\delta_{(C)}(DEPT)$	$\delta_{ m (H)} \left(J ext{ in Hz} ight)$	HMBC
1	170.3 (C)		
2	128.9 (CH)	5.93 (br. s)	H-C(5)
3	167.4 (C)		H-C(5), H-C(8')
4	42.2 (CH ₂)	2.33 (br. s)	H-C(2), H-C(5), H-C(8')
5	20.5 (Me)	2.00 (br. s)	H-C(2)
1′	42.4 (C)		H-C(6')
2′	198.6 (C)		H-C(4)
3′	115.0 (CH)	5.74 (br. <i>s</i>)	$H_a - C(5'), H_\beta - C(5'), H - C(10')$
4′	162.3 (C)		$H_a - C(5'), H - C(10')$
5'	36.1 (CH ₂)	2.29 (dd , $J = 13.2, 5.7, H_a$), 2.37 (dd , $J = 13.2, 9.9, H_\beta$)	$H-C(3'), H_{\beta}-C(7'), H-C(10')$
6′	34.0 (CH)	1.88 (<i>m</i>)	
7′	34.5 (CH ₂)	1.73 (br. $d, J = 10.8, H_{a}$), 1.79 ($t, J = 10.8, H_{\beta}$)	H-C(4), H-C(8'), H-C(9')
8′	19.6 (Me)	1.09 (br. s)	
9′	15.7 (Me)	1.01 (d, J = 6.3)	
10′	19.9 (Me)	2.22 (br. s)	H-C(3')

be anchored at the quaternary C-atom (C(1')) in subunit **C**. As anticipated, this was also in accordance with the HMBC correlation of H-C(4) to C(2'), and the observed NOE effect between H-C(4) and H-C(8'). Regarding the geometrical configuration of the C=C bonds, both 2,3-dihydro and 3',4'-dihydro olefins were shown to be (Z)configured by the NOE enhancements discerned between H-C(2) and H-C(5), as well as between H-C(3') with H-C(10') (*Fig. 1*). In conclusion, the planar structure of metabolite **1** is 3-methyl-4-(1,4,6-trimethyl-2-oxocyclohept-3-enyl)but-2-enoic acid, which possesses a hitherto undescribed C skeleton.

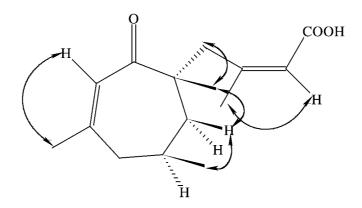


Fig. 1. Key NOED correlations of compound 1

The absolute configuration of compound **1** was established by its circular dichroism (CD) spectrum and NOE difference spectroscopy. A negative *Cotton* effect at 320 nm ($\Delta \varepsilon = -0.121$) ascribable to the $n \rightarrow \pi^*$ transition of conjugated ketone (C(2')) allowed the assignment of (1'S) for metabolite **1** [4–6]. In the NOE difference spectrum of compound **1**, pronounced NOE enhancements were observed with H_{β}-C(7') *triplet* at δ 1.79 ($J(6',7'\beta) = 10.8$ Hz) upon irradiating both H–C(8') singlet at δ 1.09 and H–C(9') doublet (J(6',9') = 6.3 Hz) at δ 1.01, allowing the assignment of the (6'R)-configuration for acid **1**.

Based on the above information, a computer-generated plot for the 3D structure of **1** (*Fig.* 2) was obtained according the process reported in [4]. The calculated distances between H–C(3')/H_a–C(10') (3.387 Å), H–C(3')/H_β–C(10') (2.345 Å), H–C(3')/H_p–C(10') (3.501 Å), H–C(2)/H_a–C(4) (3.502 Å), H–C(2)/H_β–C(4) (2.291 Å), H–C(2)/H_p–C(4) (3.281 Å), H_β–C(7')/H_a–C(8') (3.663 Å), H_β–C(7')/H_β–C(8') (2.704 Å), H_β–C(7')/H_p–C(8') (2.493 Å), H_β–C(7')/H_a–C(9') (2.990 Å), H_β–C(7')/H_β–C(9') (2.513 Å), H_β–C(7')/H_p–C(9') (3.741 Å), H_β–C(4)/H_a–C(8') (3.234 Å), H_β–C(4)/H_β–C(8') (3.867 Å), H_β–C(4)/H_p–C(8') (2.671 Å), H_a–C(4)/H_a–C(8') (3.757 Å), H_a–C(4)/H_β–C(8') (4.386 Å), and H_a–C(4)/H_p–C(10') (3.829 Å), are less than 4.00 Å, consistent with the well-defined NOEs observed for each of these H-atom pairs. Thus, the structure of **1** is (2*Z*)-3-methyl-4-[(1*S*,6*R*,3*Z*)-1,4,6-trimethyl-2-oxocyclohept-3-enyl]but-2-enoic acid.

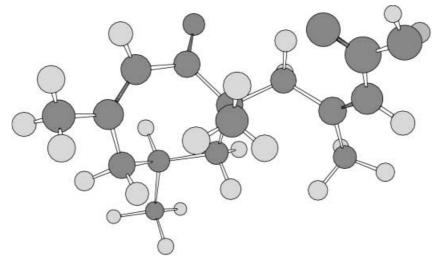


Fig. 2. 3D Structure of compound 1

Experimental Part

General. All chemicals used in the study were of anal. grade. Optical rotations: *JASCO DIP-181* spectrometer. UV Spectra: *Hitachi U-3000* spectrophotometer. CD Spectrum: *J-20C* automatic spectropolarimeter. IR Spectra: *Nexus 870 FT-IR*, ν in cm⁻¹. ¹H- and ¹³C-NMR, DEPT, HMQC, HMBC, NOED Spectra: *Bruker DPX-300* spectrometer; at 300 and 75 MHz, resp.; δ in ppm rel. to Me₄Si as an internal standard, coupling constants *J* in Hz. HR-ESI-MS (positive-ion mode): *VG-ZAB-HS* mass spectrometer.

Material. Leptosphaeria sp., strain number IV403, is an endophytic fungus isolated from fresh stems of apparently healthy *Artemisia annua* collected in May, 1997, in the suburb of Nanjing, China [7]. The re-culture broth of *Leptosphaeria* sp. was obtained according to the process reported in [4].

Extraction and Isolation. The culture filtrate (total volume 60 l) and mycelium were extracted with AcOEt. Evaporation of solvent *in vacuo* gave a residue (11.0 g), which was then subjected to chromatography on silicagel column (250 g, 200–300 mesh) eluting with petroleum ether/acetone gradient (1:0–0:1) to give five fractions (*Fr. 1:* 5.1 g; *Fr. 2:* 1.3 g; *Fr. 3:* 1.7 g; *Fr. 4:* 2.1 g; *Fr. 5:* 0.7 g). *Fr. 4* was subjected to silica-gel chromatography (60 g, 200–300 mesh) with CHCl₃/MeOH (15:1 \rightarrow 5:1). Three fractions were obtained (*Fr. 4.1:* 0.8 g; *Fr. 4.2:* 0.6 g; *Fr. 4.3:* 0.6 g). Further CC separation of *Fr. 4.2* over silica gel with CHCl₃/MeOH (8:1; 0.3 l), followed by repeated gel filtration over *Sephadex LH-20* with CHCl₃/MeOH 1:1 gave **1** (4 mg).

Leptosphaeric Acid (=(2Z)-3-*Methyl-4-[(1S*,6R,3Z)-*1*,4,6-*trimethyl-2-oxocyclohept-3-enyl]but-2-enoic Acid*, 1: white gum. [a] $_{D}^{5}$ = +63.0 (CHCl₃, c = 0.073); UV: (CHCl₃) λ_{max} 242; IR: 3500-2500 (br.), 2957.4, 2922.7, 1713.3, 1692.9, 1668.4, 1642.9, 1456.5, 1379.1, 1252.7. CD (c = 0.10 mg/ml in CHCl₃): λ 320 ($\Delta \epsilon$ - 0.121). ¹H- and ¹³C-NMR (300 and 75 MHz, resp.; CHCl₃): see *Table*. HR-ESI-MS: 251.1641 ([M+H]⁺, C₁₅H₂₃O₃⁺; calc. 251.1642).

This work was financed by grant for R.-X. T. from NNSF (No. 39970083).

REFERENCES

- [1] R. X. Tan, W. X. Zou, Nat. Prod. Rep. 2001, 18, 448.
- [2] W. X. Zou, J. C. Meng, H. Lu, G. X. Chen, G. X. Shi, T. Y. Zhang, R. X. Tan, J. Nat. Prod. 2000, 63, 1529.
- [3] H. Lu, W. X. Zou, J. C. Meng, J. Hu, R. X. Tan, *Plant Science* 2000, 151, 67.
- [4] J. Y. Liu, C. H. Liu, W. X. Zou, X. Tian, R. X. Tan, Helv. Chim. Acta 2002, 85, 2664.
- [5] A. Yajima, K. Mori, *Tetrahedron Lett.* 2000, 41, 351.
- [6] G. Snatzke. Tetrahedron 1965, 21, 43.
- [7] C. H. Liu, W. X. Zou, H. Lu, R. X. Tan, J. Biotechnol. 2001, 88, 277.

Received September 9, 2002